indicates that the complexes are formed in a 1:1 ratio. Values of the association constant, K, and of the molar extinction coefficient ϵ are reported in Table I.

The spectra of these change transfer complexes are similar in both contour and intensity to the spectrum of GPD in presence of DPN.⁶⁻⁹ This suggests that in the GPD-DPN complex the pyridinium moiety of the coenzyme molecule interacts with indole side chains of the enzyme¹⁰; this interaction should almost certainly occur in the case of GPD-APDPN complex, since iodoacetate only slightly reduces the absorption in the $360 \text{ m}\mu \text{ region.}^9$

Complexing said to be associated with electron transfer from the indole nucleus to flavines and pteridines has been reported recently.¹¹

Acknowledgment.—We are indebted to the Rockefeller Foundation and to the Brazilian "Conselho Nacional de Pesquisas" for grants, and to the latter also for a fellowship granted to P. G.

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45, 136 (1959).

DEPARTMENT OF CHEMISTRY

FACULDADE DE FILOSOFIA, CIENCIAS E LETRAS UNIVERSIDADE DE SÃO PAULO GIUSEPPE CILENTO SÃO PAULO, BRASIL PAOLA GIUSTI DEDENUED MAY 5, 1050

Received May 5, 1959

A NEW METHOD FOR THE INTRODUCTION OF THIOL GROUPS INTO PROTEINS

Sir:

For fundamental and technical reasons many attempts have been made to introduce sulfur *de novo* into proteins. Using thioglycolides, Schöberl¹ prepared highly thiolated casein and ovalbumin

$$(-SCH_{2}CO-SCH_{2}CO-)_{x} + \frac{H_{2}N}{H_{2}N} \text{protein} \longrightarrow$$
$$HS-CH_{2}CO-NH \text{protein} \quad (1)$$
$$HS-CH_{2}CO-NH \text{protein} \quad (1)$$

Since Schöberl's reagent is difficult to characterize, the recent method of Benesch and Benesch^{2,3} using N-acetylhomocysteine thiolactone is more attractive

(1) A. Schöberl, Angew. Chem., 60, 7 (1948).

R. Benesch and R. E. Benesch, THIS JOURNAL, 78, 1597 (1956).
 R. Benesch and R. E. Benesch, Proc. Natl. Acad. Sci., 44, 848 (1958).

$$H_{2}C C = 0$$

$$H_{2}C - CH$$

$$H_{2}C - CH$$

$$NHCOCH_{3}$$

$$HS - CH_{2}CH_{2}CHCO - NH - protein$$

$$NHCOCH_{3}$$

$$(2$$

Although direct reaction is not especially useful,⁴ with Ag⁺ as adjuvant³ thiolation of gelatins has been effected rapidly at pH 7.5. However, the presence of silver, and subsequently 1 M thiourea for its removal, presents problems particularly with disulfide-containing proteins.

These problems are avoided by a suitable extension of the reaction of acid anhydrides with proteins.⁵ For example with S-acetylmercaptosuccinic anhydride

$$CH_{3}CO-S-CH-C \bigcirc O + H_{2}N-protein \longrightarrow CH_{2}-C \bigcirc O + H_{2}N-protein \longrightarrow CH_{3}CO-S-CHCO-NH-protein (II) CH_{2}COOH + HS-CHCO-NH-protein (3) (3)$$

(III) CH₂COOH

Solid anhydride⁶ is added to protein solution at, for example, pH7, over from 0.25–1 hour, depending on the amount of reagent. The pH is maintained by adding sodium hydroxide. Air is excluded throughout with nitrogen. (Coupling can be performed over a range of pH and temperature.) Hydrolyzed anhydride (I) is removed with an anion exchanger, salts with a mixed-bed exchanger or by dialysis. The mercaptosuccinylated protein is isolated by lyophilization.

Typical results at room temperature are sum marized in Table I.

Mercaptosuccinvl Proteins

Protein	Moles (I) added per 10 ⁵ grams of protein	pH of reaction	Moles intr 10 ⁵ grams Mer- capto- succinyl (III)	oduced per of protein Acetyl- mer- capto- succinyl (II)
Gelatin	3 0	7	6	12
Gelatin	120	8	12	17
Gelatin	3 60	8	14	23
Bovine serum albumin	45	8	5	21
Bovine serum albumin	360	8	8	54
Ribonuclease	140	7	2	6

A reaction analogous to (3) giving mercaptosuccinylated esters has been achieved with polyhydroxylic molecules (*e.g.*, dextran, polyvinyl alcohol).

(4) It has been successfully applied at high pH (10.7) by S. J. Singer, J. E. Fothergill and J. R. Shainoff, THIS JOURNAL, **81**, 2277 (1959).

 (5) H. Frankel-Conrat, R. S. Bean and H. Lineweaver, J. Biol. Chem., 177, 385 (1949); P. H. Maurer and H. Lebovitz, J. Immunol., 76, 335 (1956); A. F. S. A. Habeeb, H. G. Cassidy and S. J. Singer, Biochim. Biophys. Acta, 29, 587 (1958).

(6) B. Holmberg and E. Schjånberg, Arkiv Kemi Mineral. Geol., 14A, No. 7 (1940).

That the mercapto groups are really bound to protein, and not merely the result of inefficient separation, was shown by ultracentrifugation with a colored azomercurial. At 60,000 r.p.m., the schlieren and color boundaries moved together.

The acetyl-S linkage of (II) is stable for days in aqueous solution at pH's as high as 9.5. Conversion of (II) into (III), if desired, can be accomplished in a few minutes in dilute sodium hydroxide, pH11.5. Preliminary experiments indicate that some nitrogenous bases also work at pH's much nearer 7 (*e.g.*, imidazole).

DEPARTMENT OF CHEMISTRY	
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RECEIVED MAY 25,	1959

HIGH ENERGY EXCHANGE REACTION OF TRITIUM ATOMS WITH CYCLOPROPANE

Sir:

Our recent experiments with tritium atoms slowing down from very high energies in the presence of cyclopropane show a substantial incorporation into the organic molecule by reaction (1), in which the asterisk designates an energetic species

$$T^{*} + \underbrace{CH_{2} - CH_{2}}_{CH_{2}} \longrightarrow C_{3}H_{6}T^{*} \longrightarrow H + \underbrace{CH_{2} - CH_{2}}_{CHT}(1)$$

Previous experiments with thermal deuterium atoms have failed to show any exchange of D for H in the cyclopropane molecule.¹

Gaseous mixtures of He⁸ and cyclopropane, with oxygen or He⁴ sometimes added, have been irradiated with thermal neutrons to produce tritium by the reaction He³(n,p)H⁸. The resulting radioactive products have been separated and measured with a proportional counter on the outlet end of a gas chromatographic column.² The percentage of radioactivity incorporated in each radioactive product is shown in Table I for several runs, both with and without added gases.

In these systems, O_2 serves as a very effective radical scavenger.³ The essentially unchanged yield of cyclopropane in its presence indicates that free radicals are not involved, and that the reaction goes through an intermediate as indicated in (1). Presumably the absence of observable exchange with thermal deuterium atoms is the result of a high activation energy for this reaction; the recoil tritium atoms react as "hot" atoms before reaching thermal energies. Moderating collisions with He³ or He⁴ serve to reduce the average energy of the tritium atom at the time of reaction,⁴ and hence reduce the possibility of exchange during collision. This is reflected in the lower yield of cyclopropane in the He⁴ experiments.

Such irradiations cause degradation of the parent molecules by ordinary radiation effects. In the 70.4 cm. Hg Δ run of Table I. the final gaseous mixture contained about 1% other hydrocarbons

(4) See, for example, M. El-Sayed, P. Estrup and R. Wolfgang, J. Phys. Chem., 62, 1356 (1958).

 TABLE I

 RADIOACTIVE PRODUCTS OF THE GASEOUS REACTION OF

 ENERGETIC TRITIUM ATOMS WITH CYCLOPROPANE

Gas Pressure, cm.	70.4∆ 2.0 He ^s	Per cent. t 31.44 1.9 He ³	otal observ 21.5Δ 1.9 He ³	ved tritium ⁶ 10.1Δ 1.5 He ³	8.4Δ 1.9 He ^a
Irradiation conditions n./ cm.²/sec. Product	$^{6 \text{ days at}}_{3 \times 10^9}$	${}^{12}_{2}~{}^{\mathrm{hr.~at}}_{10^{12}}$	$8.6 0_2$ 12 hr. at 2×10^{12}	66.7 He° 6 days at 3 \times 109	12 hr. at 12×10^{12}
Δ	22.1	16.4	15.3	10.9	7.4
HT	31.2	47.4	58.3	30.1	54.9
$CH_{3}T$	2.5	6.8	6.0	1.8	6.8
C-C	6.2	4.7	4.0	7.8	6.4
C=C	1.5	1.8	1.8	2.0	2.0
C-C-C	12.0	6.5	4.1	13.7	6.8
C-C=C	2.1	2.5	2.2	1.8	1.7
c_c_c	3.0	1.5	1.0	3.1	1.8
C-C-C-C	8.6))	13.8	1
$\sim < c$	Low	${}^{4.9}$	$^{1.8}$	Low	5.0
с с-с-с	5.4	4.0	1.5	7.8	3.7
С-С-С-С-С	1.4	1.7	0.7	1.8	0.9
}-c	0.7	0.5	0.1	0.5	<0.1
" Smaller (mounte	<107 eg	ch) hove	heen oh	erved for

^a Smaller amounts (<1% each) have been observed for $C \equiv C$, C = C = C, C = C

others.

than the parent, principally ethane and propane. Runs for higher *nvt* irradiations showed a higher percentage of radiation damage. Quantitative explanations of the distribution of radioactivity will require separation of the energetic tritium atom reactions from the accompanying macroscopic radiation damage.

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RECEIVED MAY 15, 1959		

OXAMYCIN, A COMPETITIVE ANTAGONIST OF THE INCORPORATION OF D-ALANINE INTO A URIDINE NUCLEOTIDE IN STAPHYLOCOCCUS AUREUS

Sir:

Oxamycin (D-4-amino-3-isoxazolidone, D-cycloserine), like penicillin, bacitracin, novobiocin and gentian violet, induces uridine nucleotide accumulation in *S. aureus.*¹ The nucleotides which accumulate are bacterial cell wall precursors.² Their accumulation, as well as protoplast formation,³ is the consequence of inhibition of cell wall synthesis by these antibacterial substances.

The major compound isolated from oxamycintreated cells had a slower mobility in several solvents than UDP-GNAc-lactyl-(L)ala-(D)glu-(L)lys-(D)ala-(D)ala,^{4,5} the principal compound which

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(2) J. T. Park and J. L. Strominger, Science, 125, 99 (1957).

(3) J. Lederberg, J. Bacteriol., 73, 144 (1957).

(4) In this abbreviation UDP refers to uridine diphosphate and GNAc-lactyl to an ether of acetylglucosamine and lactic acid (acetylmuramic acid). The peptide, for which the usual abbreviations are employed, is linked to the carboxyl group of the lactic acid. Its sequence recently has been determined.⁶

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